GENETIC POLYMORPHISMS AND RACIAL GROUPS IN THE POPULATION OF PINAR DEL RIO PROVINCE (CUBA)

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PALABRAS CLAVE: Marcadores genéticos, Polimorfismo, Grupos raciales, Pinar del Río, Cuba

RESUMEN: Se estudió el polimorfismo genético de 13 marcadores genéticos en tres grupos raciales (blancos, mulatos y negros) en la provincia de Pinar del Río (Cuba). Se encontraron diferencias fenotípicas en 8 de los 13 marcadores estudiados. Las diferencias de las frecuencias alélicas encontradas en los marcadores fueron altamente significativas entre los grupos raciales examinados. Los loci estudiados permitieron mostrar claras diferencias de un grupo racial respecto de los otros. Los hallazgos obtenidos permiten confirmar la hipótesis de que la población estudiada no es homogénea. Rev. Arg. Antrop. Biol. 4(1): 9-20, 2002.

KEY WORDS: Genetic markers, Polymorphism, Racial groups, Pinar del Río, Cuba

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ABSTRACT: The polymorphism of 13 genetic markers in three racial groups (whites, mulattos and blacks) of the population of Pinar del Río province (Cuba) was investigated. Differences among phenotypic rates in 8 of the 13 markers studied were found. The allelic frequencies in the markers that showed differences were highly significant among the racial groups. The loci investigated were able to clearly differentiate one racial group from the others. This finding confirms the presumption that the population of this region is not homogeneous. Rev. Arg. Antrop. Biol. 4(1): 9-20, 2002.

INTRODUCTION

The Pinar del Río province is located in the western tip of the Island of Cuba. Its population is near 697.986 inhabitants (Censo de Población y Viviendas, 1981) and occupies an area of 10.924,56 Km².

The population of this province, as well as that from Cuba, is the product of migrations of different racial groups: whites of Spanish origin, negroes from Africa and Chinese from Asia. In 1492 this province was the least populated region in Cuba.

This part of Cuba was originally inhabited by aborigines of the Guanahatabey cultural group (Carreras, 1985). The onset of Spanish colonization originated a racial mixture between whites and aborigines. Towards the end of the XVIth century, few traces of the Guanahatabey cultural group remained (Pérez de la Riva, 1972; Alonso, 1990). The Spaniards that colonized this region, during the first decade of the XVth century, were primarily from Canary Islands and Castilla (Le Riverend, 1974; Sochegui, 1980).

African Negroes were brought to Cuba as slaves since the Spaniards settlement and during colonization (XVth century) (Moreno Fraginals, 1978). The number of slaves brought to Cuba during colonization is thought to be over one million (Pérez de la Riva, 1970). The exact origin of this population is unknown, but it is believed that most of them were taken from places along the west coast of Africa (from Senegal to Angola) (Moreno, 1976; Moreno Fraginals, 1977; Rivero de la Calle, 1981). The racial admixture between whites and blacks created the "mulatto" group. This group is a typical element in the Cuban population (Hidalgo, 1986). The Chinese people were brought to Cuba in the middle of the XIXth century from Kwangtung and Fukien provinces. Despite the great number of Chinese coolies who arrived in Cuba (124.000), only 49 were females, according to the 1899 Census (Pérez de la Riva, 1967). Consequently, the Asian immigration left only a few traces on the structure of the Cuban population. This group represents a small fraction of the studied population and its possible contribution was not taken into account.

Studies of genetic markers in the Cuban population are scarce. They are limited mainly to Mas-Martín et al. (1964) (ABO and RH (D)) in Habana City (capital of Cuba), González et al. (1976), García et al. (1982) (GLO1), Barrios and Granda (1983) and Hidalgo (1986) in the central region of Cuba. In Pinar del Río province only our studies on the ABO (without A1 and A2 phenotypes) (Díaz, 1985) and HP systems (Díaz et al., 1995) have been performed.

The objective of this work is to investigate the polymorphism distribution of 13 genetic markers in the population of Pinar del Río province and to determine whether there exist phenotypic and allelic differences in each genetic marker studied among the three typical racial groups of this population: whites, mulattos and blacks.

MATERIAL AND METHODS

The sample was randomized and collected from unrelated volunteer blood donors, from Pinar del Río province Blood Banks. Among 95% donors were males and aged 20 to 40. All subjects were born in Pinar del Río, so were their parents and grandparents. The conditions of the study were communicated to all participants and each of them signed a written consent form authorizing their blood analysis.

To ensure the sample was representative of the province, the subjects were selected from each political-administrative region (municipalities) of the province.

The classification of subjects into racial groups (White, Mulatto and Black) was made according to two criteria: direct observations of morphologic characters (Hidalgo, 1986) and personal interviews where specific questions on the ancestry were asked to each of the participants by a single interviewer. We joined Mulattos and Blacks in a new group referred to as Negroids.

Blood samples were obtained by venous puncture with anticoagulant. Plasma and cells were separated by centrifugation. Then plasma was frozen at -30°C so as to study the CHE2, PI, GC and TF systems. The red blood cells were subjected to two different processes: 1) resuspension of erythrocytes, which was made to determine phenotypes of ABO (without A1 and A2), MN and P systems; 2) crioconservation, as described by Issitt (1985). This was undertaken for a later study of the HB, GLO1, PGM1 and G6PD systems, A1 and A2 phenotypes of ABO system and phenotypes of KELL and RH (haplotypes) systems.

The samples were transferred from the Pinar del Río Blood Bank to the Human Genetic Laboratory of The Higher Medical Science Institute of Santa Clara, Villa Clara (Cuba), at -30°C.

The phenotype determination of ABO (without A1 and A2), MN and P systems was made by the method described by Levine (1954). The KELL, RH (haplotypes), and A1 and A2 phenotypes were studied by the microenzymatic technique described by The American Association of Blood Banks (1985). The phenotype determination of Hemoglobin (HB), Glucose-6-Phosphate Dehydrogenase (G6PD) and Pseudocholinesterase (CHE2) systems was typed according to the methods described by Ciscar (1972), Beutler (1966) and Heredero et al. (1974), respectively; and the phenotypes of Glyoxalase I (GLO1), according to Taggart et al. (1978), and Palmour et al. (1980). The phenotypes of Alpha-1-antitripsin (PI) system were determined by the method described by Kueppers (1976). The phenotypes for the Group Specific Component (GC) and Transferrin (TF) were determined according to Dannewitz (1985), and the phenotypes of Phosphoglucomutase I (PGM1) system were obtained by the method published by Dykes and Polensky (1981).

The homogeneity of phenotype frequencies among racial groups was tested by the chi-square test of homogeneity (X^2) . The significance level was chosen at p<0.01 (negroids were excluded from this comparison).

The allelic frequency estimates were obtained by the maximum likelihood method. In systems with one completely dominant allele we assumed that the Hardy-Weinberg equilibrium was reached. In the estimation of haplotype frequencies of the RH system five sera were employed: anti-C, anti-C, anti-D, anti-E and anti-e, considering the simultaneous presence of Cde and CDE.

The Hardy-Weinberg equilibrium condition was calculated according to standard procedures described by Smith (1970). A p<0,01 level of significance with one degree of freedom was used. For those loci with more than two alleles, the expected genotypic proportions were obtained by means of an obvious extension of the Hardy-Weinberg law. In those cases, the degrees of freedom were calculated as the difference between the number of alleles and the number of possible phenotypes (Workman et al., 1963).

RESULTS

The results of phenotypic and allelic frequencies of all genetic systems are presented in Tables 1 to 4 for whites, mulattos, blacks and negroids in general, respectively.

The X² test for homogeneity of phenotypes among racial groups was highly significant (p<0,01) in the ABO, P, RH (haplotypes), TF, GC, G6PD, HB and GLO1 (Table 5).

The X^2 (H-W) test was not significant for all the racial groups and (for all the) genetics markers (Tables 1 to 4).

DISCUSSION

If all genes with a frequency higher than 0,01 are considered polymorphic (Harris, 1980), it can be stated, according to our study, that hemoglobin is not polymorphic in the white and black groups. Moreover, certain systems such as CHE2 and TF show polymorphic borderline frequencies.

The differences observed in the allelic frequencies between the different racial groups are important. Thus, the Pinar del Río population is not homogeneous. These findings are in agreement with previous results obtained by González et al. (1976); Hidalgo (1986); García et al. (1982); Barrios and Granda (1983) in other Cuban populations.

Upon analyzing the heterogeneity between the different racial groups it was observed among the 13 systems that most of them showed significant differences. If data about HP (haptoblobin) system obtained by Díaz et al. (1995) in the same population are included, we can see that 9 out of 14 genetic systems in that population showed highly significant differences. This racial heterogeneity has been maintained by more than 17 generations since the Spaniard's arrival to Cuban Islands. Possibly, assortative mating maintains these groups apart.

The non-significant deviation from the Hardy-Weinberg equilibrium obtained by the goodness-of-fit test between the observed and the expected phenotypic frequencies suggests that the conditions of random mating, homogeneity of subsamples, and absence of selection are roughly fulfilled in each group.

These findings confirm our hypothesis about the existence of differences between racial groups in Pinar del Río. Taken together, these loci discriminate well among the racial groups examined here (64,4% of these polymorphisms studied in our population showed significant differences).

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Phenotypic and allelic frequencies in each genetic system studied in Whites Table 1

9		1 1 1 1 1 1			1		
ABO		NELL		GLOI	I		11
A1	50	KK 0		1-1 27	MM 126		1-1 104
A2	20	Kk 26		2-1 80	MS 27		2-2 6
В	9	kk 281		2-2 48	O SS		3-3 0
A1B	4	n 307		n 155	7	0,9117±1,6214E-2	1-3 3
A2B	0	KEL*K	$0,0423\pm0,00812$	KEL*K 0,0423±0,00812 GLO1*1 0,4322±0,0281	S*Id	0,0883±1,6214E-2	2-3 1
0	72	KEL*k	0.9577 ± 0.00812	KEL*k 0,9577±0,00812 GLO1*2 0,5657±0,0281	31 $\chi 2 = 1,37; 1 \text{ d.f.}; p>0,01$	l.f.; p>0,01	1-2 39
n l	152	$\chi 2 = 0.66$	$\chi 2 = 0.60$; 1 d.f.; p>0.01	$\chi 2 = 0.36$; 1 d.f.; p>0,01		•	n 153
ABO*A1	ABO*A1 0,1962±0,02412		•				TF*C1 0,8169±2,2104E-2
ABO*A2	ABO*A2 0,0859±0,01842	RH		G6PD	PGM1		TF*C2 0,1699±2,147E-2
ABO*B	ABO*B 0,0333±0,01038	CCDEe	1	Normals 145	1A 62		TF*C3 0,0132±6,493E-2
ABO*O	$0,6844\pm0,02902$	CCDee	30	Deficients 3	1B 6		$\chi 2 = 1,08; 3 \text{ d.f.}; p>0,01$
$\chi 2 = 3,19;$	$\chi 2 = 3,19; 2 \text{ d.f.}; p>0,01$	CcDEe	15	G6PD*+ 0,9797±0,0082	82 1A1B 17		
		CcDee	58	G6PD*- 0,0203±0,0082	82 2A 7		GC
MN		Ccddee	0	n 148	2B 1		1F 4
MM 52		ccDEE	4	HB	2A2B 3		1S 59
		ccDEe	17	AA 305	1A2A 39		2-2 8
NN 32		ccDee	9	AS 3	1A2B 8		2-1F 1
n 181		ccddee	20	AC 0	1B2A 7		2-1S 57
MN*M	MN*M 0,5552±2,6118E-2 ccddEe	ccddEe	_	n 308	1B2B 3		1F-1S 24
WN*N	MN*N 0,4448±2,6118E-2 n	u	152	HB*A 0,9951±7,91E-6	n 153		n 153
$\chi 2 = 1,30;$	$\chi 2 = 1,30; 1 \text{ d.f.}; p>0,01$	Haplotypes	sec	HB*S 0,0049±7,91E-6		PGM*1A 0,6144±0,0278	GC*1F 0,1078±1,7731E-2
		CDe 0,4	CDe 0,43906±0,02867	$\chi 2 = 0.007$; 1 d.f.; p>0.01	. ,	PGM*1B 0,1274±0,0190	GC*1S 0,6503±2,7260E-2
Ь		Cde 0,0	$0,00000\pm0,00003$		PGM1*2A 0,2059±0,0231	$2059\pm0,0231$	GC*2 0,2418±0,2447
P1 136		CDE 0,0	$0,00501\pm0,00532$	CHE2	PGM*2B 0,0523±0,0123	$0523\pm0,0123$	$\chi 2 = 11,42; 3 \text{ d.f.}; p>0,01$
P2 16		cDE 0,	$0,12495\pm0,02027$	C5+ 13	$\chi 2 = 9,10$; 6 d.f.; p>0,01	.f.; p>0,01	•
n 152		cdE 0,	$0,00819\pm0,00822$	C5- 140			
P*1 0,67	$0,6755\pm0,0383$	cDe 0,	$0,05194\pm0,01978$	n 153			
P*2 0,32	P*2 0,3245±0,0383	cde 0,	$0,37279\pm0,02773$	CHE2*- 0,9566±0,0116	2		
		$\chi 2 = 1,81$,81; 3 d.f.; p>0,01	CHE2*+ 0,0434±0,0116	2		

 χ 2: Chi-Square of Hardy-Weinberg equilibrium

Phenotypic and allelic frequencies in each genetic system studied in Mulattos Table 2

TF	1-1 93 7-2 1	0 2-2	64E-2 1-3 0	64E-2 2-3 0	0,01 1-2 17	n 111	TF*C1 0,9144±1,8775E-2	TF*C2 0,1699±1,8775E-2	$\chi 2 = 8,03E-2; 1 \text{ d.f.}; p>0,01$			25	1F 34	15 19	2-2 1	2-1F 9	2-1S 3	1F-1S 45	n 111	0,0315 GC*IF 0,5495±3,3392E-2	E0,0233 GC*1S 0,3873±3,2695E-2	-0,0242 GC*2 0,0630±1,6314E-2	PGM*2B 0,0357±0,0123 χ 2=2,46;3d.f.;p>0,01	,01				
PI MM 101	MM 101 MS 0	SS 1	PI*M 0.9505±1.4564E-2		$\chi 2 = 2,47$; 1 d.f.; p>0,01	n 1111		PGM1	1A 50	1B 3	1A1B 20		2B 0	2A2B 1	1A2A 23	1A2B 6	1B2A 5	1B2B 1	n 112	PGM*1A 0,6652±0,0315	PGM*1B 0,1429±0,0233 GC*1S	PGM1*2A 0,1562±0,0242 GC*2	PGM*2B 0,0357±	$\chi 2 = 0.63; 6 \text{ d.f.}; p > 0.01$				
GLO1	1-1 10 2 1 64			GLO1*1 0,3750±0,0320	KEL*k 0,9505±0,01029 GLO1*2 0,6250±0,0320	$\chi 2 = 3,11; 1 \text{ d.f.}; p>0,01$	•	G6PD	Normals 95	Deficients 9		G6PD*- 0,0865±0,0195	n 104	HB	AA 217	AS 6	AC 0	n 223	HB*A 0,9865±5,45E-3	HB*S 0,0134±5,45E-3	$\chi 2 = 0.035; 1 \text{ d.f.}; p>0.01$		CHE2	C5+ 4	C5- 107	n 111	CHE2*- 0,9818±0,0089	CHE2*+ 0,0182±0,0089
KELL	KK 1	NA 20 14 201	n 222	KEL*K 0,0495±0,01029 GLO1*1 0,3750±0,0320	KEL*k 0,9505±0,01029	$\chi 2 = 0.42$; 1 d.f.; p>0.01		RH	CCDEe 1	CCDee 15	CeDEe 10	CcDee 32	Ccddee 0	ccDEE 1	ccDEe 20	ccDee 23	ccddee 7	ccddEe 1	n 110	Haplotypes	CDe 0,32810±0,03223	Cde 0,00000±0,00005	CDE 0,00826±0,00863	cDE 0,13213±0,02681	cdE 0,01415±0,01521	cDe 0,26731±0,04528	cde 0,25003±0,04485	$\chi 2 = 4,27; 3 \text{ d.f.}; p>0,01$
ABO	A1 30	A2 13	Δ / V	A2B 0	0 56	n 110	ABO*A1 0,1576±0,02569	ABO*A2 0,0852±0,02104 RH	ABO*B 0,0418±0,01364 CCDEe	ABO*O 0,7152±0,03260	$\chi 2 = 1,05; 2 \text{ d.f.}; p>0,01$		MN	MM 42	MN 81	NN 30	n 141	MN*M 0,5392±2,8495E-2 ccddEe	MN*N 0,4608±2,8495E-2	$\chi 2 = 0.65$; 1 d.f.; p>0.01		۵.	P1 77	P2 33	n 110	P*1 0,4523±0,0398	P*2 0,5477±0,0398	

 χ 2: Chi-Square of Hardy-Weinberg equilibrium

 Table 3

 Phenotypic and allelic frequencies in each genetic system studied in Blacks

ABO	KELL		GL01	핕		TF	
A1 19	KK 2		1.1 7	MM	<u>∞</u>	1-1	
,	7 17		70				
A2 11	NK 23			MS	,	7-7	
B 21	kk 186		2-2 51		0	3-3	
AIB 0	п 213		90I u	PI*M	0.9667 ± 0.0123	1-3	
A2B 0	KEL*K 0,0681±0,0122	$1\pm0,0122$	GLO1*1 0,2924±0,0312	PI*S	$0,0333\pm0,0123$	2-3 (
0 56	KEL*k 0,9319±0,0122	9±0,0122	GLO1*2 0,7075±0,0312	$\chi 2 = 0,1$	$\chi 2 = 0.10$; 1 d.f.; p>0.01	1-2	
n 107	$\gamma 2 = 1.19$; 1 d.f.; p>0.01	. p>0.01	$\chi 2 = 0.84$; 1 d.f.; p>0.01	п 105	5	п 153	
ABO*A1 0,0936±0,02042						TF*C1	0.9715 ± 0.0114
ABO*A2 0,0596±0,01728 RH	3 RH		G6PD	PGM1		TF*C2	TF*C2 0,0285±0,0114
ABO*B 0,1045±0,02151	CCDEe 1		Normals 88	Υ	42	$\chi 2 = 7,73$	$\chi 2 = 7,73E-2; 1 \text{ d.f.}; p>0.01$
ABO*O 0,7420±0,03121	_		Deficients 15	13	2		
$\chi 2 = 4,52; 2 \text{ d.f.}; p>0,01$	CcDEe 3		G6PD*+ 0,8544±0,0246	1A1B	23		
•	CcDee 15		G6PD*- 0,1456±0,0246	2A	4	ည	
MN	Ccddee 1		n 103	2B	0	1F 4	6
MM 40	ccDEE 1		HB	2A2B	1	1S	2
MN 67	ceDEe 28		AA 195	1A2A	21	2-2	2
NN 34	ccDee 49		AS 15	1A2B	7	2-1F	9
n 141	ccddee 6		AC 1	1B2A	4	2-1S	,
MN*M 0,5213±0,0297	ccddEe 0		n 211	1B2B	1	1F-1S 40	0
MN*N 0,4787±0,0297	п 107		HB*A 0,9620±9,29E-3	-	105	n 105	ν.
$\chi 2 = 0.32$; 1 d.f.; p>0.01	Haplotypes		HB*S 0,0355±9,01E-3	PGM*1		GC*1F	$0,6857\pm3,2034E-2$
	CDe 0,09911±0,02631	0,02631	HB*C 0,0023±2,36E-3	PGM*1B	3 0,1524±0,0248	GC*1S	0,2523±2,9974E-2
Ь	Cde 0,01888±	$0,01888\pm0,01803$	$\chi 2 = 0.32; 1 \text{ d.f.}; p > 0.01$	PGM1*∵	PGM1*2A 0,1619±0,0254	GC*2	0,0619±1,6629E-2
P1 78	CDE 0,00816±0,00009	6000000	CHE2	PGM*2]	PGM*2B 0,0428±0,0139	$\chi 2 = 8,12$	$\chi 2 = 8,12; 3 \text{ d.f.}; p>0,01$
P2 29	cDE 0,15071±0,02533	-0,02533	C5+ 3	$\chi 2 = 1.9$	$\chi 2 = 1,97; 6 \text{ d.f.}; p>0,01$		
n 107	cdE 0,00000±0,00004	-0,00004	C5- 103				
P*1 0,4795±0,0412	cDe 0,48289±	$0,48289\pm0,05009$	n 105				
P*2 0,5205±0,0412	cde 0,24023±	0,24023±0,04666	CHE2*- 0,9856±0,0082				
	$\chi 2 = 7,96; 3 \text{ d.f.; p} > 0,01$:p>0,01	CHE2*+ 0,0144±0,0082				

 χ 2: Chi-Square of Hardy-Weinberg equilibrium

 Table 4

 Phenotypic and allelic frequencies in each genetic system studied in Negroids in general

ABO	KELL		GL01	PI	TE
A1 49	KK 3		1-1 17	MM 199	1-1 192
A2 26	Kk 45		2-1 112		2-2 1
	kk 387		2-2 93		3-3 0
AIB 2	n 435		n 218	PI*M 0,9583±0,0096	1-3 0
A2B 0	KEL*K	KEL*K 0,0586±0,0079	GLO1*1 0,3288±0,0222	PI*S 0,0417±0,0096	2-3 0
0 112	KEL*k	KEL*k 0,9414±0,0079	GLO1*2 0,6712±0,0222	$\chi 2 = 1,27; 1 \text{ d.f.}; p>0,01$	1-2 23
n 217	$\chi 2 = 1.71$	$\chi 2 = 1.71$; 1 d.f.; p>0.01	$\chi 2 = 4.39$; 1 d.f.; p>0.01	n 216	n 216
ABO*A1 0,1257±0,01647		•			TF*C1 0,9421±0,0112
ABO*A2 0,0721±0,01356	5 RH		G6PD	PGM1	TF*C2 0,0579±0,0112
ABO*B 0,0721±0,01265	CCDE	2	Normals 183	1A 92	$\chi 2 = 0.12$; 1 d.f.; p>0.01
ABO*O 0,7299±0,02247	7 CCDee	18	Deficients 24	1B 5	•
$\chi 2 = 4,23; 2 \text{ d.f.}; p>0,01$	CcDEe	13	G6PD*+ 0,8841±0,0157	1A1B 43	
•	CcDee	47	G6PD* 0,1159±0,0157	2A 7	GC GC
MN	Ccddee	1	n 207	2B 0	1F 83
MM 82	ccDEE	2	HB	2	15 24
MN 148	ccDEe	48	AA 412	1A2A 44	2-2 3
NN 64	ccDee	72	AS 21	13	2-1F 15
n 294	ccddee	13	AC 1	6	2-1S 6
MN*M 0,5306±2,0580E-2	2 ccddEe	_	n 434	2	1F-1S 85
MN*N 0,4693±2,0580E-2	2 n	217	HB*A 0,9746±5,34E-3	n 217	n 216
χ 2 = 0,03; 1 d.f.; p>0,01	Haplotypes	sec	HB*S 0,0242±1,17E-3	PGM*1A 0,6544±0,0228	GC*1F 0,6158±0,0234
	CDe 0,2	CDe 0,21319±0,02186	HB*C 0,0012±1,17E-3	PGM*1B 0,1475±0,0170	GC*1S 0,3218±0,0224
Δ.	Cde 0,0	Cde 0,01063±0,00954	$\chi 2 = 0.28; 1 \text{ d.f.}; p > 0.01$	PGM1*2A 0,1590±0,0175	GC*2 0,0625±0,0116
PI 155	CDE 0,0	CDE 0,00889±0,00685	CHE2	PGM*2B 0,0391±0,0093	$\chi 2 = 6,67; 3 \text{ d.f.}; p>0,01$
P2 62	cDE 0,1	$0,14037\pm0,01869$	C5+ 7	$\chi 2 = 1,57;6 \text{ d.f.}; p>0,01$	
n 217	cdE 0,0	0,00741±0,00800	C5- 209		
P*1 0,4655±0,0286	cDe 0,3	$0,37665\pm0,03437$	n 216		
P*2 0,5345±0,0286	cde 0,2	$0,24283\pm0,03253$	CHE2*- 0,9837±0,0061		
	$\chi 2 = 13.9$	$\chi 2 = 13,94; 3 \text{ d.f.; p} > 0,01$	CHE2*+ 0,0163±0,0061		
				-	

 Table 5

 Chi-Square of homogeneity among whites, mulattos and blacks in each genetic system studied

System	χ2	d.f.	р	
ABO	27,15	8	<0,001	
MN	2,36	4	>0,05	
KELL	4,52	4	>0,05	
P	17,74	2	<0,0001	
RH	101,61	18	<0,0001	
CHE2	4,93	2	>0,05	
PI	11,48	4	>0,01	
TF	31,85	8	<0,001	
HB	15,46	2	<0,0005	
G6PD	13,81	2	<0,005	
GC	181,21	10	<0,0001	
GLO1	13,74	4	<0,01	
PGM1	10,90	18	>0,05	
HP	15,15	4	<0,01	(Díaz et al., 1995)

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